

Nutrient uptake and yield of rice (*Oryza sativa*) applied with mycorrhizal fungi using different doses of nitrogen and phosphorus fertilizers

ACHMAD FATCHUL AZIEZ*

Department of Agrotechnology, Faculty of Agriculture
Universitas Tunas Pembangunan, Surakarta, Central Java 57135, Indonesia
*(e-mail : achmad.aziez@lecture.utp.ac.id)

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ABSTRACT

Rainfed rice fields in general often lack water and nutrients that are difficult for their roots to reach. The Vesicular Arbuscular Mycorrhiza (VAM) fungus can assist in absorbing water and nutrients to increase the efficiency of nitrogen and phosphorus fertilizers. The purpose of this study was to determine the uptake of N, P, and K nutrients and the yield of rainfed lowland rice applied with VAM at different doses of N and P. This study was conducted in rainfed rice fields in Demangan, Central Java, Indonesia, during 2019. This research was laid out in completely randomized block design with two factors and three replications. The first factor was nitrogen with a dose of 0, 45, 90, and 135 kg/ha, while the second factor was phosphorus at 0, 25, 50, and 75 kg/ha. The results showed that VAM could increase the uptake of nitrogen, phosphorus, and potassium and increase yield using N and P fertilizers at 90 kg/ha and 75 kg/ha, respectively. This research shows that the use of N and P fertilizers at 90 kg/ha and 75 kg/ha, respectively, is sufficient to cultivate rainfed rice applied with mycorrhizal fungi. Based on the discussion, it can be concluded that the application of mycorrhizae is beneficial in rainfed rice cultivation because it can suppress the use of N and P fertilizers by increasing the uptake of N, P and K nutrients and increasing yields. In addition, in rice cultivation in rainfed rice fields, it is better to use mycorrhizae.

Key words : Growth, nitrogen, phosphorus, rainfed lowland rice, vesicular arbuscular mycorrhizae

INTRODUCTION

Rainfed land uses rainwater for irrigation and differs from irrigated rice fields. Rainfed rice fields have a low available P content due to groundwater leaching (Meng *et al.*, 2018; Anusha *et al.*, 2020). In general, improper agricultural management, long-term application of chemical fertilizers, and inefficient fertilizer use decrease the soil productivity of rice fields (Durai Singh *et al.*, 2021).

Drought stress is one of the most destructive abiotic stresses affecting plant growth and development (Shultana *et al.*, 2019). Drought stress affects physiological processes, biochemical changes, formation of secondary metabolites, significantly accumulates endogenous reactive oxygen species (ROS), and increases toxin levels (Hasanuzzaman *et al.*, 2017; Nguyen and Tran, 2019).

Drought stress greatly reduces rice grain yields and vegetative growth (Ahadiyat *et al.*, 2014; Maisura *et al.*, 2014). Water-scarce conditions generally reduce grain size, grain weight, and seed formation rates (Raman *et al.*, 2012; Kumar *et al.*, 2014). Drought stress during the booting, flowering, and terminal stages can interfere with floret initiation, cause grain sterility, lower grain weight, and ultimately lower grain yield (Acuña *et al.*, 2008). The rate of grain yield loss depends on the duration of water scarcity, plant growth stage, and stress intensity (Gana, 2011; Kumar *et al.*, 2014).

One of the efforts to overcome drought stress is microbial-based technology, such as vesicular-arbuscular mycorrhizae (Carvajal and Gloria, 2020). Mycorrhizae can act as a link between plant roots and soil moisture, especially during the dry season. Plants with mycorrhizal roots showed increased nutrient

absorption than that those without mycorrhizal roots (Narwal *et al.*, 2018). Abd-Alla *et al.* (2014) stated that mycorrhizae increase nutrient uptake by expanding the absorption range through external hyphae that can reach 8 cm outside the root system, exploiting micropores as the outer diameter of small hyphae is less than 20% of the root hair diameter, and increasing the surface area of the absorption system.

Hernández and Munné-Bosch (2015) added that the application of mycorrhizae increased the absorption rate and content of phosphorus in seeds. Phosphorus is crucial in the photosynthetic cycle as it helps activate Ribulose 1,5 biphosphate carboxylase oxygenase (Rubisco) and the Calvin cycle. Mycorrhizal fungal exudates can affect P leaching in the soil to ensure the availability of P to plants and may affect the absorption of other macronutrients (Tran *et al.*, 2020). The purpose of this study was to determine the uptake of N, P, and K nutrients and the yield of rainfed lowland rice applied with VAM fungi at different doses of N and P.

MATERIALS AND METHODS

The authors conducted this study in rainfed rice fields in Demangan, Sambu, Boyolali, Central Java, Indonesia, from March 30 2019 to July 14 2019. This study used a completely randomized block design with two factors and three replications. The first factor was urea with a dose of 0, 45, 90, and 135 kg/ha, while the second factor was phosphorus at 0, 25, 50, and 75 kg/ha. The planting process used three seeds. The authors selected one plant at 14 days old and used KCl as the fertilizer with a dose of 50 kg/ha.

Before planting, each plot was evenly sown with 5 kg of mycorrhizal fungus. The

length and width of the experimental plots were 200 cm and 120 cm, respectively. The water level was 5 cm deep, with the plants spaced 20 cm × 20 cm apart. The experimental field was weeded at 2 and 4 weeks after planting and controlled pests and diseases using organic pesticides. The harvest criterion was the seed shells above the panicle being clean and firm. Observations included nitrogen, phosphorus, leaf potassium uptake, and grain weight per plot. Nutrient uptake in leaves was calculated using the formula :

$$\text{Leaf nutrient uptake} = \text{Leaf tissue nutrient content} \times \text{Leaf dry weight.}$$

Statistical analysis used the Anova SAS 9.1 program and continued with Duncan's Multiple Distance Test (DMRT) at a probability level of 5%.

RESULTS AND DISCUSSION

Leaf Nitrogen Uptake

Nutrient uptake is the ability of plants to absorb nutrients from the soil and convert them into plant parts. The higher the plant's ability to absorb nutrients, the higher the yield obtained (Wang *et al.*, 2014). The nitrogen and phosphorus fertilizers had a significant effect on nutrient uptake. Table 1 shows that without nitrogen fertilizer, there was no difference in nitrogen uptake using 45, 75 and 90 kg/ha of phosphorus fertilizer. However, 135 kg/ha of nitrogen fertilizer combined with 50 and 75 kg/ha of phosphorus fertilizer showed higher nitrogen uptake than with 0 kg/ha and 25 kg/ha of phosphorus fertilizer.

Applying N and P fertilizers with a dose of up to 45 and 75 kg/ha, respectively did not improve nitrogen uptake. Starting from 90 kg/

Table 1. Leaf nitrogen uptake of rice applied with mycorrhizal fungi using different doses of nitrogen and phosphorus fertilizers ($\mu\text{g}/\text{mm}^2/\text{sec}$).

Dose of N (kg/ha)	Dose of P (kg/ha)				Mean
	0	25	50	75	
0	1.730 g	3.037 fg	2.267 g	2.437 g	2.367
45	3.740 fg	4.433 e-g	5.200 d-g	5.130 d-g	4.625
90	7.150 c-f	8.107 c-e	8.647 cd	9.997 bc	8.475
135	10.773 bc	13.320 b	17.480 a	19.250 a	15.205
Mean	5.848	7.224	8.398	9.203	

Figures followed by the same letter show no significant difference at the 5% DMRT level.

ha of N fertilizer, a significant increase in N uptake was seen. Dakshina Murthy *et al.* (2015) also reported increased rice nutrient uptake with increased fertilizer doses. In addition to soil nutrient supply, mycorrhizal fungi also affect nutrient uptake (Bernardo *et al.*, 2017; Narwal *et al.*, 2018; Saboor *et al.*, 2021). This finding matches with the results of Cavagnaro *et al.* (2012), Lee and Muneer (2014) and Narwal *et al.* (2018) who stated that the application of mycorrhizal fungi increases nitrogen uptake and content in seeds and plants, so that it will be able to increase dry weight (Weisany *et al.*, 2021).

Leaf Phosphorus Uptake

The nitrogen and phosphorus fertilizers affect phosphorus uptake. Table 2 shows no difference in phosphorus uptake between plants application with 25, 50, and 75 kg/ha of phosphorus fertilizer and the control plants. The same applies to plants application with 45 and 90 kg/ha of nitrogen fertilizer. However, plants application with 135 kg/ha of nitrogen fertilizer showed the highest phosphorus nutrient uptake when combined with 50 kg/ha of phosphorus fertilizer. The results significantly differ from 0, 25, and 75kg/ha of phosphorus fertilizer. The highest phosphorus uptake was 5.821 g/mm²/second, achieved by applying nitrogen and phosphorus fertilizers at 135 and 50 kg/ha, respectively. The lowest phosphorus uptake was 1,150 g/mm²/second, shown in plants without N and P fertilizers.

Phosphorus uptake was not affected by the dose of P fertilizer; higher doses did not increase P uptake. Phosphorus uptake of rice plants fertilized with phosphorus was relatively low, presumably due to a high increase in fertilization which decreased nutrient uptake and the ability of rice plants to absorb

phosphorus. Continuous application of phosphate fertilizers causes P accumulation, reducing plant response to phosphate fertilization. This fact matches the findings of (Liu *et al.*, 2020). Besides reducing P efficiency, P accumulation can also affect the availability of other nutrients. In plants, P is an integral part of its cellular activities. Phosphorus is crucial in plant metabolism processes, such as cell division, development, photosynthesis, sugar breakdown, nutrient transport, and metabolic pathway regulation (Bagyaraj *et al.*, 2015). Arbuscular mycorrhizal fungi can increase the efficiency of P fertilizer absorption. This fungus can achieve symbiosis with roots and is crucial in ecological and agronomical plant growth. According to Bolduc and Hijri (2011), nutrient uptake in plants with mycorrhizal roots is more efficient than plants without mycorrhizae. This condition is due to the absorption and transport of nutrients by mycorrhizae.

Abd-Alla *et al.* (2014) stated that mycorrhizae increase nutrient uptake by expanding the absorption range through external hyphae that can reach 8 cm outside the root system, exploiting micropores as the outer diameter of small hyphae is less than 20% of the root hair diameter, and increasing the surface area of the absorption system. Plants with mycorrhizae usually perform better than those without mycorrhizae. Mycorrhizae can effectively increase the absorption rate of macronutrients (N, P, K, Ca, Mg, and Fe) and micronutrients (Cu, Mn, and Zn) of plant root hairs. This condition improves plant metabolism, indicated by increased plant crown growth (Dakshina Murthy *et al.*, 2015).

Ortas (2015) stated that the dry weight and P uptake of corn plants without mycorrhizae would increase along with the increase in the dose of SP-36. The higher the

Table 2. Leaf phosphorus uptake of rice applied with mycorrhizal fungi using different doses of nitrogen and phosphorus fertilizers ($\mu\text{g}/\text{mm}^2/\text{sec}$).

Dose of N (kg/ha)	Dose of P (kg/ha)				Mean
	0	25	50	75	
0	1.150 f	1.992 d-f	1.707 ef	1.305 f	1.538
45	2.095 d-f	2.482 c-f	3.384 b-d	1.828 d-f	2.447
90	2.172 d-f	3.040 b-e	3.942 bc	2.686 b-f	2.960
135	2.709 b-f	3.714 bc	5.821 a	4.040 b	4.071
Mean	2.031	2.807	3.713	2.464	

Figures followed by the same letter show no significant difference at the 5% DMRT level.

P concentration in the soil, the larger the role of mycorrhizae in absorbing P. Nazirah *et al.* (2018) added that mycorrhizae are crucial for plants as they increase phosphorus uptake and drought resistance. Upland plants, including Situbagendit, have a positive response to mycorrhizae.

Leaf Potassium Uptake

The nitrogen and phosphorus fertilizers affect potassium nutrient uptake. Without the nitrogen fertilizer, phosphorus fertilizer with a dose of 0 to 75 kg/ha did not improve potassium uptake. The same is true for nitrogen fertilization at 45 and 90 kg/ha. However, 135 kg/ha of nitrogen fertilizer combined with 50 kg/ha of phosphorus fertilizer provided the highest potassium uptake and showed different results from 0 kg/ha and 25 kg/ha of phosphorus fertilizer (Table 3). Nitrogen plays a dominant role in potassium absorption, while mycorrhizae do not. Various references state that mycorrhiza plays a role in nitrogen and phosphorus absorption, but its role in potassium absorption is still unclear. Researchers have thoroughly studied the effects of mycorrhizal symbiosis for nitrogen and phosphorus uptake (Plassard and Dell 2010). However, few have studied the possible effects of mycorrhizae on potassium uptake

(Ruíz-Sánchez *et al.*, 2011). Baslam *et al.* (2011) stated that the K content increased in rice plants application with mycorrhizae.

Grain yield

The nitrogen and phosphorus fertilizers do not affect grain yield. The application of nitrogen fertilizer at 0, 45, 90, and 135 kg/ha showed the same results (Table 4). Plants applied with mycorrhizae only require 45-90 kg/ha of nitrogen fertilizer. Bahadur *et al.* (2019) stated that mycorrhizae can help the host plant absorb the nutrients needed for photosynthesis, while the host photosynthesizes for the mycorrhizal fungi. The inoculation of mycorrhizal fungi can increase the yield of soybeans, peanuts, green beans, corn, and sweet potatoes. Ebrahim and Saleem (2017), Saboor *et al.* (2021) and Okonji *et al.* (2018) stated that utilizing mycorrhizae as biological fertilizers containing microorganisms greatly reduces nutrients absorbed by colloidal soils due to low pH or Al and Fe activity. In the long term, application of mycorrhizae will greatly benefit soil fertility.

CONCLUSION

Based on the discussion, it can be concluded that the application of mycorrhizae

Table 3. Leaf potassium uptake of rice applied with mycorrhizal fungi using different doses of nitrogen and phosphorus fertilizers ($\mu\text{g}/\text{mm}^2/\text{sec}$).

Dose of N (kg/ha)	Dose of P (kg/ha)				Mean
	0	25	50	75	
0	6.33 e	8.83 c-e	7.10 de	10.19 c-e	8.11
45	10.85 c-e	11.26 c-e	13.22 b-d	10.37 c-e	11.42
90	11.38 c-e	11.83 c-e	13.54 bc	12.54 b-e	12.32
135	11.63 c-e	14.85 bc	21.32 a	18.43 ab	16.55
Mean	10.04	11.69	13.79	12.88	

Figures followed by the same letter show no significant difference at the 5% DMRT level.

Table 4. Rice grain yield per plot (200 cm x 120 cm) applied with mycorrhizal fungi using different doses of nitrogen and phosphorus fertilizers (g).

Dose of N (kg/ha)	Dose of P (kg/ha)				Mean
	0	25	50	75	
0	523.3 bc	510.0 c	786.7 a-c	556.7 a-c	594.2 b
45	767.3 a-c	701.0 a-c	768.3 a-c	768.7 a-c	751.3 ab
90	847.0 a-c	961.3 ab	729.3 a-c	926.7 a-c	866.1a
135	940.0 a-c	874.7 a-c	659.7 a-c	988.0 a	865.6 a
Mean	769.4 a	761.7 a	736.0 a	810.0 a	

Figures followed by the same letter show no significant difference at the 5% DMRT level.

is beneficial in rainfed rice cultivation because it can suppress the use of N and P fertilizers by increasing the uptake of N, P and K nutrients and increasing yields. In addition, in rice cultivation in rainfed rice fields, it is better to use mycorrhizae.

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REFERENCES

- Abd-Alla, M. H., El-Enany, A. W. E., Nafady, N. A., Khalaf, D. M. and Morsy, F. M. (2014). Synergistic interaction of *Rhizobium leguminosarum* Bv. Viciae and Arbuscular Mycorrhizal fungi as a plant growth promoting biofertilizers for faba bean (*Vicia Faba* L.) in alkaline soil. *Microbiol. Res.* **169** : 49-58. doi: 10.1016/j.micres.2013.07.007.
- Acuña, T. L. Botwrih., Lafitte, H. R. and Wade, L. J. (2008). Genotype × environment interactions for grain yield of upland rice backcross lines in diverse hydrological environments. *Field Crops Res.* **108** :117-25. doi: 10.1016/j.fcr.2008.04.003.
- Ahadiyat, Y. R., Hidayat, P. and Susanto, U. (2014). Drought tolerance, phosphorus efficiency and yield characters of upland rice lines. *Emirates J. Food Agric.* **26** : 25-34. doi: 10.9755/ejfa.v26i1.14417.
- Anusha, P. L., Umamahesh, V., Ramana Rao, P. V., Rama Rao, G. and Subba Rao, M. (2020). Physiological and biochemical characterization of rice (*Oryza sativa*) genotypes suitable for dry direct sowing condition. *Crop Res.* **55** : 189-201.
- Bagyaraj, D. J., Sharma, M. P. and Maiti, D. (2015). Phosphorus nutrition of crops through arbuscular mycorrhizal fungi. *Curr. Sci.* **108** : 1288-293. doi: 10.18520/cs/v108/i7/1288-1293.
- Bahadur, A., Batool, A., Nasir, F., Jiang, S., Mingsen, Q., Zhang, Q., Pan, J., Liu, Y. and Feng, H. (2019). Mechanistic insights into arbuscular mycorrhizal fungi-mediated drought stress tolerance in plants. *Int. J. Mol. Sciences* **20** : 1-18. doi: 10.3390/ijms20174199.
- Baslam, M., Garmendia, I. and Goicoechea, N. (2011). Arbuscular mycorrhizal fungi (AMF) improved growth and nutritional quality of greenhouse-grown lettuce. *J. Agric. Food Chem.* **59** : 5504-515. doi: 10.1021/jf200501c.
- Bernardo, L., Morcia, C., Carletti, P., Ghizzoni, R., Badeck, F. W., Rizza, F., Lucini, L. and Terzi, V. (2017). Proteomic insight into the mitigation of wheat root drought stress by arbuscular mycorrhizae. *J. Proteom.* **169** : 21-32. doi: 10.1016/j.jprot.2017.03.024.
- Bolduc, A. R. and Hijri, M. (2011). The use of mycorrhizae to enhance phosphorus uptake: A way out the phosphorus crisis. *J. Biofert. Biopest.* **02** : 1-5. doi: 10.4172/2155-6202.1000104.
- Carvajal, M. and Gloria, B. (2020). Genetic regulation of water and nutrient transport in water stress tolerance in roots. **324** : 134-42. doi: 10.1016/j.jbiotec.2020.10.003.
- Cavagnaro, T. R., Barrios-Masias, F. H. and Jackson, L. E. (2012). Arbuscular mycorrhizas and their role in plant growth, nitrogen interception and soil gas efflux in an organic production system. *Plant Soil* **353** :181-94. doi: 10.1007/s11104-011-1021-6.
- Dakshina Murthy, K. M., Upendra Rao, A., Vijay, D. and Sridhar, T. V. (2015). Effect of levels of nitrogen, phosphorus and potassium on performance of rice. *Indian J. Agric. Res.* **49** : 83-87. doi: 10.5958/0976-058X.2015.00012.8.
- Durai Singh, R., Hussainy, S. A. H., Paulpandi, V. K., Nandhini, R., Lavanya, A. and Prema, M. (2021). Effect of integrated nutrient management on the growth, phyllochron, tillering and yield of rice (*Oryza sativa*). *Crop Res.* **56** : 281-86.
- Ebrahim, M. K. H. and Saleem, A. (2017). Alleviating salt stress in tomato inoculated with mycorrhizae : photosynthetic performance and enzymatic antioxidants. *Integ. Med. Res.* **11** : 850-60. doi: 10.1016/j.jtusci.2017.02.002.
- Gana, A. (2011). Screening and resistance of traditional and improved cultivars of rice to drought stress at Badeggi, Niger State, Nigeria. *Agric. Biol. J. North America* **2** : 1027-31. doi: 10.5251/abjna.2011.2.6.1027.1031.
- Hasanuzzaman, M., Nahar, K., Bhuiyan, T. F., Anee, T. S., Inafuku, M., Oku, H. and Fujita, M. (2017). Salicylic acid: An all-rounder in regulating abiotic stress responses in plants. In: Phytohormones - signaling mechanisms and crosstalk in plant development and stress responses. pp. 31-74.
- Hernández, I. and Munné-Bosch, S. (2015). Linking phosphorus availability with photo-

- oxidative stress in plants. *J. Exptl. Bot.* **66** : 2889-900. doi: 10.1093/jxb/erv056.
- Kumar, S., Dwivedi, S. K., Singh, S. S., Bhatt, B. P., Mehta, P., Elanchezhian, R., Singh, V. P. and Singh. (2014). Morpho-physiological traits associated with reproductive stage drought tolerance of rice (*Oryza Sativa* L) genotypes under rain-fed condition of eastern Indo-Gangetic plain. *Indian J. Plant Physiol.* **19** : 87-93 doi: 10.1007/s40502-014-0075-x.
- Lee, B. R. and Muneer, S. (2014). Mycorrhizal colonisation and P-supplement effects on N uptake and N assimilation in perennial ryegrass under well-watered and drought-stressed conditions. *Mycorrhiza* **22** : 525-34. doi: 10.1007/s005 doi: 10.1007/s00572-012-0430-6.
- Liu, M., Zhao, Z., Chen, L., Wang, L., Ji, L. and Xiao, Y. (2020). Ecotoxicology and environmental safety in Fl Uences of arbuscular mycorrhizae, phosphorus fertiliser and biochar on alfalfa growth, nutrient status and cadmium uptake. *Ecotoxicol. Environ. Safety* **196** : doi: 10.1016/j.ecoenv.2020.110537.
- Maisura, M. Chozin, I. Lubis, A. Junaedi and Ehara, H. (2014). Some physiological character responses of rice under drought conditions in a paddy system. *J. ISSAAS* **20** :104-14.
- Meng, C., Liu, H., Wang, Yi., Li, Y., Zhou, Ji., Zhou, P., Liu, X., Li, Y. and Wu, J. (2018). Response of regional agricultural soil phosphorus status to net anthropogenic phosphorus input (NAPI) determined by soil pH value and organic matter content in subtropical China. *Chemosphere* **200** : 487-94. doi: 10.1016/j.chemosphere.2018.02.125.
- Narwal, E., Annapurna, K., Choudhary, J. and Sangwan, S. (2018). Effect of arbuscular mycorrhizal fungal colonization on nutrient uptake in rice aerobic conditions. *Int. J. Curr. Microbiol. Appl. Sci.* **7** :1072-093. doi: 10.20546/ijcmas.2018.704.118.
- Nazirah, L., Purba, E., Hanum, C. and Rauf, A. (2018). Effect of soil tillage and mycorrhiza application on growth and yields of upland rice in drought condition. *Asian J. Agric. Biol.* **6** : 251-58.
- Nguyen, H.L. and Tran, D. H. (2019). Performance of salt tolerant rice cultivars under different soil salinity levels in Central Vietnam. *Res. Crop.* **20** : 461-67.
- Okonji, C., Sakariyawo, O., Okeleye, K., Osunbiyi, A. and Ajayi, E. (2018). Effects of arbuscular mycorrhizal fungal inoculation on soil properties and yield of selected rice varieties. *J. Agric. Sci., Belgrade* **63** :153-70. doi: 10.2298/jas1802153o.
- Ortas, I. (2015). Comparative analyses of turkey agricultural soils : Potential communities of indigenous and exotic mycorrhiza species ' Effect on maize (*Zea Mays* L) growth and nutrient uptakes. *European J. Soil Biol.* **69** : 79-87. doi: 10.1016/j.ejsobi.2015.05.006.
- Plassard, C. and Dell, B. (2010). Phosphorus nutrition of mycorrhizal trees. *Tree Physiol.* **30** : 1129-139. doi: 10.1093/treephys/tpq063
- Raman, A., Verulkar, S. B., Mandal, N. P., Variar, M., Shukla, V. D., Dwivedi, J. L., Singh, B. N., Singh, O. N., Padmini Swain, Ashutosh K. Mall, S. Robin, R. Chandrababu, Abhinav Jain, Tilatoo Ram, Shailaja Hittalmani, Stephan Haefele, Hans Peter Piepho and Arvind Kumar (2012). Drought yield index to select high yielding rice lines under different drought stress severities. *Rice* **5** : 1-12. doi: 10.1186/1939-8433-5-31.
- Ruiz-Sánchez, M., Armada, E., Muñoz, Y., Salamone, I. E. G., Aroca, R., Ruiz-Lozano, J. M. and Rosario Azcón (2011). Azospirillum and arbuscular mycorrhizal colonization enhance rice growth and physiological traits under well-watered and drought conditions. *J. Plant Physiol.* **168** : 1031-037. doi: 10.1016/j.jplph.2010.12.019.
- Saboor, A., Ali, M. A., Hussain, S., El Enshasy, H. A., Hussain, S., Ahmed, N., Gafur, A., Sayyed, R. Z., Fahad, S., Danish, S. and Datta, R. (2021). Zinc nutrition and arbuscular mycorrhizal symbiosis effects on maize (*Zea Mays* L.) growth and productivity. *Saudi J. Biol. Sci.* **28** : 6339-351. doi: 10.1016/j.sjbs.2021.06.096.
- Shultana, R., Othman, R., Zuan, A. T. K. and Yusop, M. R. (2019). Evaluation of growth and nutrient uptake of rice genotypes under different levels of salinity. *Res. Crop.* **20** : 1-9.
- Tran, Cuc T. K., Stephanie J. Watts-Williams, Ronald J. Smernik and Timothy R. Cavagnaro (2020). Effects of plant roots and arbuscular mycorrhizas on soil phosphorus leaching. *Sci. Total Environ.* **722** :137847. doi: 10.1016/j.scitotenv.2020.137847.
- Wang, W., Sardans, J., Zeng, C., Zhong, C., Li, Y. and Peñuelas, J. (2014). Responses of soil nutrient concentrations and stoichiometry to different human land uses in a subtropical tidal wetland. *Geoderma* **232-234** : 459-70. doi: 10.1016/j.geoderma.2014.06.004.
- Weisany, W., Tahir, N. A. and Schenk, P. M. (2021). Coriander/soybean intercropping and mycorrhizae application lead to overyielding and changes in essential oil profiles. *European J. Agron.* **126** : doi: 10.1016/j.eja.2021.126283.